Maternal obesity and the human milk metabolome: associations with infant body composition and postnatal weight gain

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ABSTRACT

Background: Maternal obesity is a risk factor for childhood obesity; this is a major public health concern given that ~40% of pregnant women are either overweight or obese. Whether differences in milk composition in lean compared with obese women contribute to childhood obesity is unclear.

Objectives: We aimed to analyze relationships between maternal obesity and human milk metabolites, infant body composition, and postnatal weight gain.

Methods: This was a prospective study in which mothers intending to breastfeed exclusively, and their newborn infants, were enrolled at delivery (n = 35 mother–infant pairs). We excluded mothers with diabetes, other medical conditions, or pregnancy complications. Participants were grouped by maternal prepregnancy BMI <25 (lean) or ≥25 kg/m² (overweight/obese). We analyzed infant body composition by dual-energy X-ray absorptiometry and used untargeted liquid chromatography–gas chromatography–mass spectrometry to measure the milk content of 275 metabolites at 1 and 6 mo postpartum.

Results: At 1 mo postpartum, 10 metabolites differed between overweight/obese and lean groups with nominal P < 0.05, but none was altered with a false discovery rate <0.25. Many differentially abundant metabolites belonged to the same chemical class; e.g., 4/10 metabolites were nucleotide derivatives, and 3/10 were human milk oligosaccharides. Milk adenine correlated positively with both maternal and infant weight, these point to potential milk-dependent mechanisms for mother–child transmission of obesity. This trial was registered at NCT02535637. Am J Clin Nutr 2019;00:1–10.

Introduction

Childhood obesity has reached unprecedented levels in the United States and worldwide (1–3). Maternal obesity is one of the strongest predictors of childhood obesity, increasing risk more than 2-fold (4). Approximately 40% of women in North America are overweight or obese (ov-ob) at pregnancy onset (5), and maternal obesity accounts for 12–15% of the population attributable risk of childhood obesity (6, 7). While the mechanisms by which obesity risk is transmitted from mother to child are not fully understood, differences in infant feeding may play a role. Indeed, maternal obesity is linked to lower initiation and shorter duration of breastfeeding (8), and obesity risk is highest among non-breastfed infants of obese mothers (9). Thus, breastfeeding promotion has been a major focus of public health efforts to curb childhood obesity (10).

While breastfeeding positively impacts many aspects of maternal and child health (11–13), the literature on offspring obesity this study was funded by R01 HD080444 and R00 HD064793 from NICHD, MINECO, Graetz Foundation, Mead Johnson Nutrition, and Abbott Nutrition. The nongovernmental funders were not involved in the design, implementation, analysis, or interpretation of the study.

Supplemental Tables 1–6 and Supplemental Figure 1 are available from the “Supplementary data” link in the online posting of the article and from the same link in the online table of contents at https://academic.oup.com/ajcn/.

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Abbreviations used: 1,5-AG, 1,5-anhydroglucitol; DXA, dual-energy X-ray absorptiometry; EE, effect estimate; FDR, false discovery rate; GPE, glycerylphosphorylethanolamine; HMO, human milk oligosaccharides; KEGG, Kyoto Encyclopedia of Genes and Genomes; ov-ob, overweight or obese; WFA, weight for age.

Received August 24, 2018. Accepted for publication October 24, 2018. First published online 0, 2019; doi: https://doi.org/10.1093/ajcn/nqy334.
risk is inconclusive. A large meta-analysis of breastfeeding compared with formula feeding found only a small protective effect on adult BMI (14). When confounders such as socioeconomic status, maternal BMI, and maternal smoking are controlled, associations between duration and exclusivity of breastfeeding and later obesity are attenuated or abolished (15). Some studies have even suggested increased obesity risk: in a randomized controlled trial of breastfeeding promotion in Belarus, the intervention group had a slightly higher BMI in adolescence (16).

One possible reason for the lack of clear benefit of breastfeeding on obesity risk is that human milk might not play a mechanistic role in childhood obesity. Another possibility is that milk composition may vary according to maternal weight status (17, 18). For example, milk from obese mothers might contain differential amounts of “obesogenic” compared with “protective” constituents. Substantial evidence indicates that total fat, saturated fatty acids, insulin, leptin, TNF-α, and adiponectin are increased in milk from obese mothers; some studies also have suggested that differences in milk ω-3 and ω-6 (n-3, n-6) fatty acids, and in microbiome diversity, may be associated with maternal weight status (19–25). Despite intriguing recent studies suggesting that milk insulin and leptin may alter the development of the infant microbiome (26), it is not clear whether such differences in composition contribute to differences in weight gain in infants of obese women. For example, human milk insulin content is positively associated with maternal BMI, but inversely correlated with infant BMI; similar associations have been reported for leptin and IL-6 (22). Thus, it remains an open question whether differences in milk composition contribute to mother–child transmission of obesity.

With the current study, we used a metabolomics approach to comprehensively analyze metabolites, nutrients, and small molecules in human milk that differ according to both maternal and infant weight status. We prospectively analyzed infant body composition, in parallel with maternal milk composition, at 1 and 6 mo postpartum. We hypothesized that human milk content of individual metabolites, and groups of metabolites belonging to specific pathways, would be associated with maternal and infant weight status.

Methods

Mother–infant participants and study procedures

All research procedures and protocols were approved by the hospital committee for human subjects research and were conducted in accordance with the Helsinki Declaration. The original purpose of this pilot study was to assess the cross-sectional associations of appetite-regulating hormones and growth factors (leptin, insulin, glucose) and inflammatory factors (IL-6 and TNF-α) in human breast milk with infant size, adiposity, and lean tissue at 1 and 6 mo of age in healthy term infants in women across a broad range of BMI; analysis of the milk metabolome was done as a secondary analysis. We enrolled 31 mother–infant pairs from Oklahoma University Health Sciences Center. Eligibility criteria included intent to exclusively breastfeed from 0 to 6 mo of age and willingness to provide expressed milk samples. We excluded mothers with gestational or pregestational diabetes and/or any medical conditions in the mother or child with the potential to influence weight gain. Participants were grouped according to maternal prepregnancy BMI, with BMI <25 kg/m² defined as “lean,” and ≥25 kg/m² defined as overweight/obese (“ov-ob”). Clinical information was obtained from medical records and questionnaires. Milk was collected at 1 and 6 mo. Mother–infant pairs reported to the study site at 1 and 6 mo (±5 d) postpartum, between 08:00 and 10:00, >1.5 h since the last infant feeding, and with the mother fasted >1 h. Upon arrival, a pre-feeding infant weight was obtained using a high-sensitivity scale (Seca 728). Mothers then breastfed the infant ad libitum from both breasts. The study visit continued, including dual-energy X-ray absorptiometry (DXA), anthropometrics, questionnaires, etc., for 2–2.5 h. After the above, based on their discretion, mothers chose the breast they felt could provide the most complete milk expression (though the right was encouraged). The mother was encouraged to completely empty the entire contents of a single breast using an electric hospital-grade breast pump (Symphony, Medela, Inc.). Milk was collected into 150-mL bisphenol A-free polypropylene containers (Medela, Inc.), mixed, and centrifuged at 3000 × g for 15 min at 4°C. The fat layer was skimmed from the top using a microspatula and discarded. Samples were mixed, centrifuged, and scraped 2 more times. The aqueous phase of the sample was transferred to a new microcentrifuge tube, taking care to avoid any remaining lipid and pelleted material. Skimmed milk samples were divided into aliquots and stored at −80°C until analysis.

Infant body composition analysis

At 1 and 6 mo, we measured infant weight using a Seca 728 scale and length (crown-to-heel) with a Seca 416 infantometer (Seca). We estimated infant adiposity (relative percentage of fat, i.e., %fat), total fat mass, total lean mass, and central obesity (trunk fat mass) by DXA (Lunar scanner, GE Healthcare), as described (22, 27). To minimize variability, the same investigator (DAF) positioned the infants and performed all the DXA scans.

Metabolomic analysis

We performed untargeted metabolomics analysis using liquid chromatography–gas chromatography–mass spectrometry, as described (28–30) (Metabolon, Inc.). Two solutions of isotopically labeled reference standards were spiked at a constant level into every experimental and quality control sample, either at the beginning or at the end of the extraction process. These compounds were carefully chosen for each data stream, such that they did not interfere with measurement of endogenous compounds, and were used to monitor assay performance in every batch. Recovery standards, added at the beginning of the extraction, allowed us to monitor extraction efficiency and reproducibility. In addition, internal standards, prepared in the reconstitution solution and added in the final step, allowed for chromatographic peak alignment and were used to monitor instrument performance and data quality over the course of the run. These internal standards included: deuterated d7-glucose, d3-leucine, d8-phenylalanine, and d5-tryptophan (Cambridge Isotope Laboratories); d5-hippuric acid, d5-indole acetic acid, and d9-progesterone (C/D/N Isotopes, Inc.); bromophenylalanine (Sigma-Aldrich); and amitriptyline (MP Biomedicals, LLC).
To confirm chromatographic peak identities, all metabolite identities were checked against a reference library. We assessed process variability from the beginning to the end of the run in technical replicate samples comprising a small aliquot of every experimental sample within the study. This sample was injected periodically throughout the run. Upon measuring metabolite peaks emerging from these samples, we calculated 11% variance across all the metabolites measured.

The assay detected a total of 275 metabolites including 223 known metabolites from diverse chemical classes, including amino acids, lipids, carbohydrates, vitamins, cofactors, etc., and 52 metabolites of unknown identity (X-11616, X-11618, etc.). Metabolites undetectable in >50% of the samples were excluded from analysis and undetectable values replaced by half the minimum value detected, using Metaboanalyst 3.0 (31, 32).

We tested whether any KEGG (Kyoto Encyclopedia of Genes and Genomes) metabolic pathways were over-represented among the differentially regulated metabolites through the pathway analysis modality of Metaboanalyst 3.0, using a list of detected metabolites as the reference group.

Statistical analysis

We initially enrolled N = 37 mother–infant pairs. We excluded n = 4 due to missing data on weight gain during pregnancy, n = 1 due to insufficient milk, and n = 1 due to missing metabolite data; thus, n = 31 (16 ov-ob, 15 lean) were included in the analysis of human milk at 1 mo. We excluded n = 6 due to missing measures at 6 mo; thus, n = 26 (14 ov-ob, 12 lean) were included in the analysis of human milk at 6 mo (see Study Flowchart in Supplemental Figure 1). We calculated group means, SDs, and other descriptive statistics using JMP Pro v13 (SAS). To identify metabolites that differ in ov-ob compared with lean mothers, we calculated fold changes for each metabolite and performed two-sided unpaired Student’s t-tests. To account for multiple hypothesis testing, we calculated false discovery rate (FDR)-adjusted P values using the Benjamini–Hochberg procedure (33, 34), with Metaboanalyst 3.0. We used an FDR threshold of 0.25, which corresponded to a nominal P value of <0.01 for the human milk metabolite data set at 6 mo. (No metabolites reached this significance threshold at 1 mo.) We also calculated Pearson correlation coefficients between levels of individual metabolites and clinical characteristics including maternal prepregnancy BMI, infant weight for age percentile, and infant % fat. We used standard least-squares linear regression to adjust for potential confounders, including infant sex, gestational age, and parity, using the Fit Model function in JMP Pro.

Results

Clinical characteristics of mother–infant pairs

Demographic and clinical information for the mother–infant pairs is presented in Table 1. As expected, women in the ov-ob group had a significantly higher prepregnancy BMI than the lean group (30.9 ± 5.1 kg/m², compared with 21.7 ± 1.9 kg/m², P < 0.0001). Women in the ov-ob group were less likely to be primiparous (12.5% compared with 46.7%, P = 0.05; parity 2.9 ± 1.7 compared with 2.0 ± 1.1, P = 0.11). Race and ethnicity were similar in the 2 groups. The ov-ob group had a higher BMI at the 1-mo postpartum visit (30.9 ± 3.6 kg/m², compared with 23.5 ± 2.4 kg/m², P < 0.0001) and greater triceps skinfold thickness at 1 mo (31.4 ± 10.1 mm, compared with 19.1 ± 7.3 mm, P = 0.002). Gestational age, birth weight, and birth weight percentile (for gestational age and sex) were similar in infants of ov-ob compared with lean mothers.

Infant weight gain and body composition

At 1 mo postpartum, infants of overweight-obese mothers tended to have a higher weight for age (WFA) percentile (66% ± 23%, compared with 47% ± 29%, P = 0.06). Infants in the ov-ob group had a significantly higher absolute fat mass (DXA) (1.3 ± 0.3 kg, compared with 1.1 ± 0.3 kg, P = 0.02) and trends for higher adiposity, expressed as percentage of body weight (25.0% ± 2.5%, compared with 23.1% ± 3.2%, P = 0.07), and truncal fat (0.45 ± 0.11 kg, compared with 0.37 ± 0.14 kg, P = 0.09) (Table 1) at 1 mo. By 6 mo, however, differences in WFA percentile and body composition were attenuated.

Associations between maternal obesity and milk metabolites at 1 mo postpartum

At 1 mo postpartum, 10 human milk metabolites differed between the ov-ob and lean groups (P < 0.05, Table 2). None of these metabolites survived adjustment for multiple comparisons (FDR < 0.25). However, many of the top-ranking metabolites belonged to the same chemical class. For example, 3 of the 10 top-ranking differentially abundant metabolites were human milk oligosaccharides, and 4 were purine and pyrimidine derivatives (Figure 1 and Table 2). Pathway analysis indicated that metabolites related to pyrimidine metabolism were over-represented among the 10 differentially abundant metabolites (P = 0.035), as compared with the reference group of 168 detected metabolites with KEGG pathway annotations.

Given that mothers in the ov-ob group had a higher parity than controls, and that gestational age and infant sex can influence milk composition (36–38), we adjusted for these potential confounders using linear regression analysis. Three human milk oligosaccharides, 2-fucosyllactose [effect estimate (EE): −0.066 ± 0.030, P = 0.03], lacto-N-fucopentaose II/III (EE: 0.068 ± 0.027, P = 0.02), and lacto-N-fucopentaose I (EE: −0.072 ± 0.027, P = 0.01), remained significantly associated with maternal BMI after adjustment, as did the pure and pyrimidine derivatives orotate (EE: −0.084 ± 0.029, P = 0.008) and adenine (EE: 0.068 ± 0.030, P = 0.03, Supplemental Table 1). To test whether infant birth weight, which was numerically higher in infants ov-ob mothers, was related to the associations between maternal weight status and milk composition, we further adjusted for birth weight and noted that associations between maternal BMI and human milk oligosaccharides, orotate, and 1,5-anhydroglucitol persisted after birth weight adjustment, whereas the association between maternal BMI and milk adenine content was attenuated.

Associations between maternal BMI and milk metabolites at 6 mo postpartum

Analysis of human milk at 6 mo revealed 20 metabolites differing significantly between the ov-ob and lean groups (P < 0.05), Table 3; 6 of these survived adjustment for
TABLE 1  Demographic and clinical characteristics of mothers and infants

<table>
<thead>
<tr>
<th>Clinical and demographic characteristics</th>
<th>Lean (n = 15)</th>
<th>Overweight-obese (n = 16)</th>
<th>P value^2</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Mothers</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age, y</td>
<td>27.5 ± 5.7 (15)</td>
<td>30.5 ± 4.7 (16)</td>
<td>0.12</td>
</tr>
<tr>
<td>Prepregnancy BMI, kg/m^2</td>
<td>21.7 ± 1.9 (15)</td>
<td>30.9 ± 5.1 (16)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Parity</td>
<td>2.0 ± 1.1 (15)</td>
<td>2.9 ± 1.7 (16)</td>
<td>0.11</td>
</tr>
<tr>
<td>Race, % white</td>
<td>73.3 (15)</td>
<td>75 (16)</td>
<td>1</td>
</tr>
<tr>
<td>Gestational weight gain, kg</td>
<td>15.1 ± 6.5 (13)</td>
<td>11.9 ± 7.0 (14)</td>
<td>0.2</td>
</tr>
<tr>
<td>Excessive gestational weight gain^3, %</td>
<td>33.3 (13)</td>
<td>50 (14)</td>
<td>0.47</td>
</tr>
<tr>
<td>Postpartum BMI at 1 mo, kg/m^2</td>
<td>23.5 ± 2.4 (15)</td>
<td>30.9 ± 3.6 (16)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Triceps skinfold thickness at 1 mo postpartum, mm</td>
<td>19.1 ± 7.3 (15)</td>
<td>31.4 ± 10.1 (13)</td>
<td>0.02</td>
</tr>
<tr>
<td><strong>Infants</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sex, % males</td>
<td>66.7 (15)</td>
<td>50 (16)</td>
<td>0.47</td>
</tr>
<tr>
<td>Birth weight, kg</td>
<td>3.45 ± 0.50 (15)</td>
<td>3.68 ± 0.46 (16)</td>
<td>0.2</td>
</tr>
<tr>
<td>Birth weight percentile</td>
<td>39.5 ± 1.0 (15)</td>
<td>40.0 ± 1.1 (16)</td>
<td>0.17</td>
</tr>
<tr>
<td>Weight for age percentile, 1 mo</td>
<td>47 ± 29 (15)</td>
<td>66 ± 23 (16)</td>
<td>0.06</td>
</tr>
<tr>
<td>Body composition (dual-energy X-ray absorptiometry), 1 mo</td>
<td>1.1 ± 0.3 (15)</td>
<td>1.3 ± 0.3 (16)</td>
<td>0.02</td>
</tr>
<tr>
<td>Total fat mass, kg</td>
<td>23.1 ± 3.2 (15)</td>
<td>25.0 ± 2.5 (16)</td>
<td>0.07</td>
</tr>
<tr>
<td>Trunk fat mass, kg</td>
<td>0.37 ± 0.14 (15)</td>
<td>0.45 ± 0.11 (16)</td>
<td>0.09</td>
</tr>
<tr>
<td>Weight for age percentile, 6 mo</td>
<td>32.3 ± 30.3 (12)</td>
<td>45.5 ± 38.4 (14)</td>
<td>0.34</td>
</tr>
<tr>
<td>Body composition (dual-energy X-ray absorptiometry), 6 mo</td>
<td>32.0 ± 3.5 (12)</td>
<td>32.6 ± 4.0 (14)</td>
<td>0.67</td>
</tr>
<tr>
<td>Total fat, %</td>
<td>2.38 ± 0.55 (12)</td>
<td>2.50 ± 0.74 (14)</td>
<td>0.63</td>
</tr>
<tr>
<td>Trunk fat mass, kg</td>
<td>0.77 ± 0.29 (12)</td>
<td>0.82 ± 0.36 (14)</td>
<td>0.7</td>
</tr>
</tbody>
</table>

^1Continuous data are presented as means ± SDs (n); categorical variables are presented as a percentage (n).

^2P value calculated using Student’s 2-sided t-test for continuous data; Fisher’s exact test used for categorical data.

^3“Excessive gestational weight gain” refers to the percentage of participants exceeding 2009 Institute of Medicine (35) guidelines for gestational weight gain.

Multiple comparisons (FDR < 0.25). The differentially abundant metabolites belonged to similar chemical classes. For example, several acylcarnitines involved in branched chain amino acid metabolism were increased in human milk from ov-ob mothers, including hydroxyisovaleroylcarnitine, propionylcarnitine, and 2-methylbutyroylcarnitine (increased 123%, 45%, and 128%, respectively (P < 0.05), Figure 2A, Supplemental Table 2). Several sugar alcohols and monosaccharides were more abundant in milk from ov-ob mothers, including 1,5-anhydroglucitol, arabitol, arabinose, glucose-6-phosphate, erythritol, and ribitol (Figure 2B). Moreover, several amino acids, including ornithine, glutamine, and asparagine, were reduced by 17%, 32%, and 49%, respectively (P < 0.05), as were levels of the tryptophan metabolite kynurenate (reduced 39%, P = 0.036, Figure 2C). Pathway analysis (Metaboanalyst) indicated that metabolites belonging to the “Citrate cycle,” “Glyoxylate/dicarboxylate metabolism,” and “Alanine, aspartate and glutamate metabolism” pathways were significantly over-represented (Supplemental Table 2) among the list of differentially regulated metabolites at age 6 mo.

TABLE 2  Top-ranking differentially regulated human milk metabolites at 1 mo postpartum

<table>
<thead>
<tr>
<th>Name</th>
<th>Class</th>
<th>Fold-change overweight or obese/lean</th>
<th>P value</th>
<th>False discovery rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lacto-N-fucopentaose I</td>
<td>Oligosaccharide</td>
<td>0.383</td>
<td>0.007</td>
<td>0.86</td>
</tr>
<tr>
<td>Lacto-N-fucopentaose II or III</td>
<td>Oligosaccharide</td>
<td>1.674</td>
<td>0.04</td>
<td>0.86</td>
</tr>
<tr>
<td>2-Fucosyllactose</td>
<td>Oligosaccharide</td>
<td>0.619</td>
<td>0.04</td>
<td>0.86</td>
</tr>
<tr>
<td>AMP</td>
<td>Nucleosides, nucleotides, and analogues</td>
<td>1.761</td>
<td>0.02</td>
<td>0.86</td>
</tr>
<tr>
<td>Cyclic adenosine 3’,5’-monophosphate</td>
<td>Nucleosides, nucleotides, and analogues</td>
<td>0.433</td>
<td>0.03</td>
<td>0.86</td>
</tr>
<tr>
<td>Cytidine</td>
<td>Nucleosides, nucleotides, and analogues</td>
<td>1.335</td>
<td>0.04</td>
<td>0.86</td>
</tr>
<tr>
<td>Orotate</td>
<td>Pyrimidines and pyrimidine derivatives</td>
<td>0.704</td>
<td>0.02</td>
<td>0.86</td>
</tr>
<tr>
<td>Piperine</td>
<td>Alkaloids and derivatives</td>
<td>2.277</td>
<td>0.02</td>
<td>0.86</td>
</tr>
<tr>
<td>2-Aminobutyrate</td>
<td>Carboxylic acids and derivatives</td>
<td>1.547</td>
<td>0.03</td>
<td>0.86</td>
</tr>
<tr>
<td>X-19659</td>
<td>Unknown</td>
<td>0.510</td>
<td>0.03</td>
<td>0.86</td>
</tr>
</tbody>
</table>

^1Fold-change refers to mean abundance in overweight or obese/lean. The table includes only metabolites with P < 0.05 (2-sided t-test).

N = 15–16/group.
Maternal BMI, milk metabolome, and infant obesity

These metabolic signatures largely persisted in a linear regression analysis of associations between maternal pre-pregnancy BMI and milk metabolite abundance at 6 mo postpartum, adjusting for maternal age and infant sex; further adjustment for birth weight attenuated associations with glutamine, arabinose, and ribitol (Supplemental Table 3). Interestingly, abundance of 1,5-anhydroglucitol (1,5-AG), which was not previously described in human milk, was positively associated with maternal BMI at both 1 mo (EE: 0.089 ± 0.025, P = 0.002) and 6 mo (EE: 0.108 ± 0.028, P = 0.001), with or without adjustment for birth weight.

Overlap between milk metabolites associated with maternal BMI and those associated with infant obesity

We next asked whether human milk metabolites associated with maternal weight status were also associated with infant weight status. We looked for overlap between milk metabolites correlated with maternal BMI (Pearson correlation, P < 0.05, Supplemental Table 1) and milk metabolites correlated with infant weight status at 1 mo, assessed by infant WFA percentile, adjusting for infant sex, gestational age, and parity. As shown in Figure 3A, we found minimal overlap between the 2 sets of metabolites, with the exception of adenine, which was positively correlated with both infant weight and with maternal BMI, and the unidentified metabolite X-19656, which was negatively correlated with both infant weight and maternal weight. We next examined the overlap between milk metabolites associated with infant adiposity (% body fat assessed by DXA) at 1 mo (Supplemental Table 4) and metabolites associated with mother’s BMI. The metabolite 5-methylthioadenosine was noted to be positively correlated with both maternal BMI and infant adiposity (Figure 3B).

By 6 mo of age, we found even less overlap between the sets of metabolites correlating with maternal BMI and those correlating with infant WFA and adiposity (Supplemental Table 5). No metabolites were correlated with both maternal BMI and infant WFA at 6 mo or with maternal BMI and infant adiposity.

Associations between milk metabolites and fat accrual in infancy

Because weight gain and fat accrual during infancy are important risk factors for obesity, insulin resistance, and cardiovascular...
disease risk in later life, we next examined associations between milk metabolites and the change in fat mass from 1 to 6 mo of age, as assessed by DXA. We used linear regression to adjust for infant sex, gestational age, and maternal age. We noted a positive correlation between adenine levels in human milk at 1 mo and change in fat mass from 1 to 6 mo (EE: 1.091 ± 0.369, \( P = 0.01 \), Supplemental Table 6); as noted above, human milk adenine was also positively correlated with maternal BMI (Supplemental Table 1).

**Discussion**

In the current study, we combined detailed metabolomic analysis of human milk from lean compared with overweight or obese women, together with longitudinal analysis of weight gain and body composition in infants, to test whether differentially abundant metabolites in milk are associated with infant obesity. Previous analyses of the human milk metabolome have focused on differences in milk composition according to gestational age (39–41), postnatal age (39), and maternal health status [e.g., diet/lifestyle (42), pre-eclampsia (43), chemotherapy (44)], but our analysis is the first comprehensive analysis of the human milk metabolome in relation to both maternal and infant obesity. We demonstrate that maternal obesity is associated with modest differences in the human milk metabolome; metabolite differences did not survive adjustment for multiple corrections at 1 mo postpartum, whereas maternal obesity was modestly associated with milk metabolite content at 6 mo postpartum. We also report that levels of many human milk metabolites are associated with infant weight status and identify a subset of metabolites correlating with both maternal BMI and infant adiposity. Together, these data raise the possibility that obesity-associated differences in human milk composition might contribute to early childhood obesity, although this hypothesis would need to be confirmed in additional studies.

Breastfeeding promotion is widely advocated as a means of improving both maternal and child health outcomes, but surprisingly little is known about the bioactive compounds in human milk responsible for its beneficial health effects. Human milk contains not only nutrients, but also antibodies, cytokines, hormones, adipokines, and a distinct microbiome (45, 46) that could potentially influence metabolic outcomes in the infant. These data raise the question of whether differences in human milk composition might contribute to mother–child transmission of obesity risk. This possibility is suggested by rodent studies, in which lactational exposure to maternal obesity imparts obesity risk in the offspring (47, 48). Similarly, a study of infants of mothers fed either mother’s milk or banked human milk from nondiabetic donors found that consumption of diabetic mothers’ milk during the first week of life was associated with a higher weight at 2 y (49). Previous studies have demonstrated that milk from obese women has higher levels of insulin, leptin, TNF-\( \alpha \), and IL-6, as compared with lean women (22, 50). However, it is not clear whether these substances play a pathogenic versus protective role, or no role at all, in childhood obesity risk. If human milk composition does play a mechanistic role in childhood obesity, it is likely to be one of many additive risk factors for childhood obesity and not the sole determinant. For example, human milk may be protective as compared with formula, but milk from obese mothers might contain a lower abundance of protective factors than lean mothers’ milk. This possibility is suggested by the observations of Li et al. (9), who

### TABLE 3 Top-ranking differentially regulated human milk metabolites at 6 mo postpartum

<table>
<thead>
<tr>
<th>Name</th>
<th>Class</th>
<th>Fold-change overweight or obese/lean</th>
<th>( P ) value</th>
<th>False discovery rate</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hydroxyisovaleryl carnitine C5</td>
<td>Acylcarnitine</td>
<td>2.23</td>
<td>0.003</td>
<td>0.15</td>
</tr>
<tr>
<td>2-Methylbutyryl carnitine C5</td>
<td>Acylcarnitine</td>
<td>2.28</td>
<td>0.045</td>
<td>0.56</td>
</tr>
<tr>
<td>Propionyl carnitine C3</td>
<td>Acylcarnitine</td>
<td>1.45</td>
<td>0.02</td>
<td>0.35</td>
</tr>
<tr>
<td>Glucose 6-phosphate</td>
<td>Monosaccharide; hexose phosphate</td>
<td>2.07</td>
<td>0.01</td>
<td>0.29</td>
</tr>
<tr>
<td>1,5-Anhydroglucitol</td>
<td>Monosaccharide</td>
<td>1.37</td>
<td>0.003</td>
<td>0.15</td>
</tr>
<tr>
<td>Arabinose</td>
<td>Monosaccharide; pentose</td>
<td>1.72</td>
<td>0.01</td>
<td>0.28</td>
</tr>
<tr>
<td>Arbutol</td>
<td>Sugar alcohol</td>
<td>1.53</td>
<td>0.01</td>
<td>0.28</td>
</tr>
<tr>
<td>Erythritol</td>
<td>Sugar alcohol</td>
<td>1.25</td>
<td>0.03</td>
<td>0.45</td>
</tr>
<tr>
<td>Ribitol</td>
<td>Sugar alcohol</td>
<td>1.38</td>
<td>0.04</td>
<td>0.52</td>
</tr>
<tr>
<td>Glycerate</td>
<td>Sugar acids and derivatives</td>
<td>0.63</td>
<td>0.0005</td>
<td>0.18</td>
</tr>
<tr>
<td>Asparagine</td>
<td>Amino acid</td>
<td>0.51</td>
<td>0.01</td>
<td>0.29</td>
</tr>
<tr>
<td>Ornithine</td>
<td>Amino acid</td>
<td>0.83</td>
<td>0.002</td>
<td>0.15</td>
</tr>
<tr>
<td>Glutamine</td>
<td>Amino acid</td>
<td>0.68</td>
<td>0.002</td>
<td>0.15</td>
</tr>
<tr>
<td>Kynurenate</td>
<td>Amino acid metabolite; uinoline carboxylic acid</td>
<td>0.61</td>
<td>0.04</td>
<td>0.51</td>
</tr>
<tr>
<td>3′-4′-Hydroxyphenyllactate</td>
<td>Amino acid metabolite; phenylpropanoic acid</td>
<td>1.59</td>
<td>0.01</td>
<td>0.28</td>
</tr>
<tr>
<td>Phosphocholine</td>
<td>Phosphocholine</td>
<td>1.44</td>
<td>0.03</td>
<td>0.44</td>
</tr>
<tr>
<td>Malate</td>
<td>( \beta )-Hydroxycarboxyls and derivatives</td>
<td>1.80</td>
<td>0.002</td>
<td>0.15</td>
</tr>
<tr>
<td>Fumarate</td>
<td>Diacarboxylic acid</td>
<td>1.47</td>
<td>0.050</td>
<td>0.57</td>
</tr>
<tr>
<td>Citrate</td>
<td>Tricarboxylic acid</td>
<td>0.78</td>
<td>0.047</td>
<td>0.56</td>
</tr>
<tr>
<td>X—12,565</td>
<td>Unknown</td>
<td>1.71</td>
<td>0.02</td>
<td>0.33</td>
</tr>
</tbody>
</table>

\( N = 15–16/group. \)

"Fold-change" refers to mean abundance in overweight or obese/lean. The table includes only metabolites with \( P < 0.05 \) (2-sided \( t \)-test).
reported that the risk of childhood obesity was highest in non-
breastfed infants of obese mothers, but also demonstrated that
risk of childhood overweight was increased by maternal obesity
among breastfed infants.

We found a relatively small degree of overlap between
milk metabolites correlating with maternal BMI and those
correlating with infant adiposity. However, levels of adenine
were associated with both maternal and infant weight status,
and with fat accumulation from 1 to 6 mo of age. Moreover,
several nucleoside and nucleotide derivatives (e.g., AMP,
orotate, adenosine 3,5-cyclic monophosphate, cytidine) were
differentially abundant in milk from ov-ob compared with normal
weight women at 1 mo. Human milk has a higher abundance
of nucleotides than cow milk (51). Infant formulas are typically
supplemented with nucleotides, which may increase weight,
length, and head circumference during infancy (52). Human
milk nucleotides may also promote development of cellular
and humoral immune responses (53, 54) and differentiation
and growth of the intestinal epithelium (53, 55). More recently,
the pyrimidine nucleotide uridine has been discovered to regulate
thermogenesis during the fasting–feeding transition (56), raising
the question of whether human milk nucleotide content may
affect systemic metabolism in the infant. Our overlap analysis
also identified 5-methylthioadenosine—a sulfur-containing
nucleoside and precursor to methionine—as positively associated
both with maternal BMI and with infant fat percentage at 1 mo,
which is intriguing in light of recent reports that this metabolite
is increased in adolescents with metabolic syndrome and adults
with type 2 diabetes (57, 58).

At 1 mo postpartum, we found that maternal obesity was linked
to differences in human milk oligosaccharide (HMO) content
and composition. HMOs are a family of structurally diverse
carbohydrate polymers that represent the third most abundant
constituent in human milk (after lactose and lipids) (59). Human
milk contains >150 types of oligosaccharides, with biological
functions differing according to their structure. Distinct from
other carbohydrates, HMOs are not digested or metabolized
for energy, instead remaining intact through the small intestine.
Upon reaching the colon, HMO can be metabolized by specific
gastrointestinal bacterial species. HMOs may thus function as
“prebiotics,” selectively fueling the growth of specific bacteria
and shaping infant gut microbiome development (13). HMOs
also act as receptor decoys and block attachment of enteric
pathogens to the host, and they may protect the infant from
group B streptococcus and other neonatal infections (60). We
have previously described an association between milk content
of lacto-N-fucopentaose I and infant fat mass (61) independent
of maternal BMI. It will be essential for future studies to
test whether HMO content and composition play a causal role in infant obesity. We speculate that alterations in HMO in obese mothers may contribute to observed differences in gut microbiome composition and diversity in infants of obese mothers (24, 26).

By 6 mo postpartum, we noted that maternal obesity was associated with increased milk abundance of acylcarnitines, monosaccharides, and sugar alcohols, and reductions in amino acids and their metabolites. These patterns are similar to metabolomics signatures in plasma from individuals with obesity or type 2 diabetes: elevations in short-chain acylcarnitines (62, 63), increased hexoses and sugar alcohols [e.g., erythritol (64), glucose (65), mannose (65)], and reduced glutamine:glutamate ratio (66) have been previously reported in adults with diabetes or at risk of the disease. One of the most significant changes in human milk was the positive association between maternal BMI and milk 1,5-AG at both 1 and 6 mo postpartum. This is intriguing in light of a large body of literature demonstrating that 1,5-AG is a marker of glycemic control (67) and that reductions in plasma 1,5-AG predict incident diabetes (65); 1,5-AG has not previously been described in human milk. Taken together, these patterns raise important questions about the degree to which the maternal plasma metabolome might influence the human milk metabolome. Unfortunately, we did not assess correlations between maternal plasma and human milk metabolites, but this will be a key question for future studies.

We acknowledge the limitations of our study. First, our analysis relied on cross-sectional associations, so we cannot infer causality. Second, our sample size was relatively small, so it will be important to replicate our findings in other populations to assess generalizability. Third, we did not assess whether milk components other than metabolites (e.g., essential micronutrients, immune cells, bacteria, exosomes, cytokines, etc.) may be linked to maternal obesity. Finally, we did not obtain detailed dietary questionnaires in the participants, so it is unclear to what degree variation in dietary patterns may contribute to

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**FIGURE 3** Overlap between milk metabolites correlating with maternal weight status and those correlating with infant weight status at 1 mo postpartum. (A) Venn diagram depicting the overlap between breast milk metabolites at 1 mo postpartum that correlate with infant weight (assessed by weight for age and sex percentile at 1 mo) and metabolites correlating with maternal pre-pregnancy BMI, with all metabolites adjusted for gender, parity, and gestational age. (B) Venn diagram depicting the overlap between breast milk metabolites at 1 mo postpartum that correlate with infant adiposity (assessed by total fat % measured by dual-energy X-ray absorptiometry at 1 mo) and metabolites correlating with maternal prepregnancy BMI. All metabolites adjusted for gender, parity, and gestational age. Significant correlation defined as effect estimate with adjusted P < 0.05. (+) denotes positive correlation, and (−) denotes negative correlation. 1,5-AG, 1,5-anhydroglucitol; GPE, glycerylphosphorylethanolamine.
human milk metabolite abundance. For example, piperine, a xenobiotic metabolite found in peppers, was increased in ov-ob mothers at 1 mo, suggesting that diet may play a role. Some of the strengths of our approach include the longitudinal assessments at 1 and 6 mo and the simultaneous analysis of weight status in both mothers and infants.

In summary, we found that maternal obesity is associated with metabolomic signatures in human milk. At 1 mo postpartum, maternal BMI was modestly associated with the abundance of HMOs known to function as prebiotics, raising the possibility that obesity-associated changes in maternal milk composition may modulate infant microbiome acquisition, a hypothesis that would need to be confirmed with additional studies. By 6 mo postpartum, maternal BMI was associated with acyl carnitines, sugar alcohols, and amino acid metabolites in human milk, a pattern reminiscent of plasma metabolomic signatures in obesity and type 2 diabetes. While we found only a modest degree of overlap between human milk metabolites that correlated with maternal prepregnancy weight and those correlating with infant obesity, the identified association between milk adenine and both maternal and infant weight status raises the possibility that some milk constituents might play a pathogenic role in mother-to-child transmission of obesity.

We are grateful to Dr Mary-Elizabeth Patti for reviewing the manuscript and providing insightful suggestions.

The authors’ responsibilities were as follows—DAF: designed the study, led the clinical assessments, and reviewed/edited the manuscript; EI: interpreted and analyzed the metabolomics data and wrote the manuscript; EWD: assisted with designing the study and with statistical considerations; CL: assisted with interpretation of metabolomics data; TIM and SV: assisted with preparation of tables and figures and all authors: read and approved the final manuscript. None of the authors reports a conflict of interest related to research presented in this article.

References


